

# Effect of Pre-Harvest Application of Salicylic Acid on Strawberry Growth and Resistance Against Gray Mold Disease

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#### ABSTRACT

The present study aims to investigate the effect of salicylic acid treatment at various concentrations on strawberry (Fragaria ananassa var. Festival) growth, biochemical components, enzymes activity, and resistance to gray mold (Botrytis cinerea) during 2020-2021 growing season. Strawberry plants were planted in mid of September on raised beds in two open fields with two different types of soil (clay and sandy soil), in Qalyubia governorate-Egypt. Salicylic acid (SA), at 1, 2, 4 mM, and fenhexamid fungicide at Egyptian MOA recommended does, were applied at flowering stage. The results revealed that SA at all applied concentrations significantly improved the plant growth parameters and fruit yield compared to control or fenhexamid treatment. Maximum of plant height, root length, leaf and flower count/ plant, root and fruit fresh weight (g)/plant, and fruit volume (cm<sup>3</sup>) were recorded by SA at the rate of 4 mM, which was followed by SA at 2mM. In addition, total chlorophyll, total phenol, and antioxidant enzymes activity increased significantly by SA 4- and 2-mM application rates. Treatment of SA in vitro, or under field's condition, reduced the diminished growth of gray mold more than Fenhexamid fungicide and control treatment. We can conclude that foliar application of salicylic acid significantly increased vegetative and fruit growth of strawberry and induced the treated plant defense against gray mold disease.

**Key words:** Strawberry, Salicylic acid, Fenhexamid, gray mold, growth parameter, plant pigments, phenols, antioxidant enzymes.

#### Introduction

One of the most popular fruits in the world is the strawberry (*Fragaria ananassa* var. Festival), which is also a significant fruit grown on a large scale in Egypt. The minerals, amino acids, vitamins, and anthocyanin in strawberries are quite abundant (**Chauhan** *et al.*, **2013**). Due to their high concentrations of phenolic compounds, including anthocyanin and vitamins C, E, and -carotene, which are chemicals linked to health benefits, these are also an important source of bioactive compounds. Ellagic acid, which has a wide range of biological activities, is present in





quite high quality in it (**Meyers** *et al.*, **2003**). Because strawberries have a well-known perishable structure, their fruit is vulnerable to fungus assault. In addition, the fruit's high metabolic rate can rapidly reduce fruit quality after harvest.

The most dangerous strawberry disease is botrytis fruit rot (gray mold) and caused by the fungus *Botrytis cinerea* which is pervasive in nature. It may become dormant and stay on fruits as well as infect strawberry blooms, causing them to decay (**Raskin, 1992**). Significant yield losses are caused in Egypt by botrytis fruit rot, is the most severe and deadly disease of strawberries. When strawberry plants are not protected, frequently resulting in yield losses of up to 25% because it destroys strawberry blossoms', setting fruit, mature fruit, and leaves (**Sutton, 1990**). Gray mold is making a significant post-harvest loss factor during strawberry storage, transit, and shipment (**Sutton and Peng, 1993**). *Botrytis cinerea*, an airborne fungus, is a saprophyte, or unspecialized neurotropic fungus, that grows on rotting tissue as grayish masses of mycelium, conidiophores, and conidia (**Ciliberti** *et al.*, **2015**). According to **Kim** *et al.* (**2007**) and **Deising** *et al.* (**2008**), Botrytis has a very short incubation period and is quickly spread by wind, which makes it a major concern for all strawberry production during most of the year.

Several functions in plants, including heat production, disease resistance, seed germination, sex polarization, and ethylene synthesis, are regulated by salicylic acid (**Raskin, 1992**). Plants are vulnerable could be treated exogenously with salicylic acid in harmless amounts to increase their resistance to fungi. It also acts as a possible non-enzymatic antioxidant and plant growth regulator (**Zhang** *et al.,* **2011**). It has a high oral LD50 (rat) of 891 mg/kg and has positive impacts on human health. Therefore, SA usage is safe for human health and is expected to enhance crop quality and stress resistance (**Peng and Jiang, 2006**).

Fenhexamid is a non-systemic molecule and acts by inhibiting spore germination, germ-tube elongation, and the mycelium growth of *Botrytis cinerea* inhibits germ-tube elongation and mycelia growth (**Ha**"nßler and Pontzen, 1999)

Due to that, the current study was planned to examine the effect of pre-harvest foliar application of salicylic acid on strawberry vegetative growth, fruit quality, and resistance to gray mold disease.

#### **Materials and Methods**

#### 1. Field preparation and planting

The current study was conducted at a private farm with two well-managed experimental fields of two types of soil: A (**clay soil**) and B (**sandy soil**) in Met-Kenana village, Toukh, Qalyubia governorate - Egypt, during 2020/2021 growing season. Tables 1 and 2 provide the physical and chemical characteristics of the experimental field soil types. Strawberries (*Fragaria ananassa* var. Festival) with one well-developed crown measuring 8–10 mm in diameter was transplanted as bare-rooted, cold-stored plants. The Strawberry and Non-Traditional Crops Improvement Center of Faculty of Agriculture - Ain Shams University is where the transplants were purchased. Strawberry transplants were planted at 10<sup>th</sup> and 14<sup>th</sup> September in clay and sandy soil fields, respectively. Raised beds that were 120 cm wide, and 15-20 cm high were used to grow the





transplants with a drip irrigation system, 30 cm apart. All cultural practices of cultivation (irrigation, fertilization, weeding, and pest management) were carried out in the two experimental fields in accordance with the recommendations of the Egyptian Ministry of Agriculture (EMOA).

# 2. Design and experiment

One month after Strawberry transplanting, Salicylic acid (2-Hydroxybenzoic acid, Molecular Weight equal 138.12g/mol., purity: 99%, Sigma-Aldrich),was dissolved in dimethyl sulphoxide (DMSO), then distilled water was added for preparing concentrations of 1, 2, and 4 milli Mole (mM). Fenhexamid SC 50% fungicide was applied according to Egyptian MOA's recommendation at the rate of 600cm<sup>3</sup>/Fed to compare as a standard course of treatment. Salicylic acid treatments were performed at flowering stage, and repeated twice by 10 days intervals; meanwhile, Fenhexamid was treated once at the flowering stage. The treatments were laid out in Randomized Block design with four treatments which replicated thrice. All treatments were applied with a knapsack sprayer in the early morning. The control plants received identical doses of water and dimethyl sulphoxide at the appointed times. Since it was noted that rapid and efficient absorption by the lower leaf surface occurred, both the upper and lower leaf surfaces were sprayed until moist (**Hull et al., 1975**). The plot area was 6.0 m<sup>2</sup> included 50 plants.

#### **3.** Fruit quality and plant growth of strawberry

At the flowering stage, ten plants were randomly selected from each of the two inner rows of each experimental plot. The length of plant roots, leaves number and flowers per plant. Fresh fruit weight, and fruit volume were recorded after 90 days from transplantation.

# 4. Chemical determinations

After ten days from each treatment, some strawberry plants were randomly selected from each experiment plot, then the leaves were separated and divided into two parts: the first part was kept as fresh leaves (for pigments and enzyme activity determinations), and the second part was dried at 60°C for two days in an electric oven, Fisher Isotemp., Senior model, then ground and stored in a deep freezer.

# 4.1. Pigment Contents

A test tube contains 5 ml of DMSO and 10 mg of strawberry fresh leaves tissue in fraction were added together. Without grinding, chlorophyll and carotenoids were extracted into DMSO solvent by incubation at 65°C overnight. For the determination of chlorophyll, absorbance was measured by spectrophotometer (Shanghai Lab-Spectrum Instrument Co., Ltd Model, Alpha-1102) at 644 and 662 nm for chlorophyll A and B, and 470 nm for carotenoids as specified by **Hiscox and Israelstam (1979)**. The **Arnon equation (1949)** was used to compute total chlorophyll (Chl. a+b), chlorophyll a (Chl. a), and chlorophyll b (Chl. b), whereas **Cañal** *et al.* (**1985**) was used to calculate carotenoids.





Arnon equation: Chl. a =  $12.7 \times O.D \ 662 - 2.69 \times O.D \ 644 \ mg/l$ Chl. b =  $22.9 \times O.D \ 644 - 4.68 \times O.D \ 662 \ mg/l$ Chl. a+b =  $20.2 \times O.D \ 644 + 8.02 \times O.D \ 662 \ mg/l$ Cañal equation:

Carotenoids= A470 – 1.28 (Chl. a mg/l) + 56.7 (Chl. b mg/l) mg/l 256 X 0.906

#### 4.2. Total Phenol contents

The Folin-Ciocalteau method was used to determine the total amount of phenolic compounds in the extracts of dried strawberry leaves for all treatments (1, 2, 4 mM of SA, Fenhexamid and control) according to **El-falleh** *et al.* (2012). The total phenolic compound in the extracts of dried strawberry leaves were determined and represented as mg/ml of extract equivalent Gallic acid.

#### 4.3. Total proteins

Total proteins in the extracts of fresh strawberry leaves for all treatments (1, 2, 4 mM of SA, Fenhexamid and control) were determined by using **Bradford** method (**I976**).

#### 4.4. Enzymes activity

According to Ni *et al.* (2015), the enzyme activity was tested in the extracts of fresh strawberry leaves for all treatments (1, 2, 4 mM of SA, Fenhexamid and control).

#### 4.3.1. Catalase activity

In accordance with **Aebi 1984**, a bio diagnostic kit (No., CA 2517), was used for catalase activity assessment in the extracts of fresh strawberry leaves for all treatments (1, 2, 4 mM of SA, Fenhexamid and control).

#### 4.3.2. Phenol oxidase activity

By the method of **Ishaaya** (**I971**), phenol oxidase activity was assessed in the extracts of fresh strawberry leaves for all treatments (1, 2, 4 mM of SA, Fenhexamid and control).

#### 4.4.3. Peroxidase activity

According to **Hammerschmidt** *et al.* (**1982**), the peroxidase activity of the extracts of fresh strawberry leaves of all treatments (1, 2, 4 mM of SA, Fenhexamid and control) were determined. The change in absorbance/min/g of protein was used to express enzyme activity.

#### 5. Evaluation of fungal diseases

#### 5.1. A fungal pathogen's source.

Strawberry fruit samples that had naturally deteriorated and displayed a range of rot signs were gathered. According to **Hamouda** *et al.* (2016) infected strawberry fruit samples from each field were sent to the laboratory.





#### 5.2. Isolation and purification of the fungus

Strawberry fruits for each treatment (SA, fenhexamid, and control) were washed with distilled water, dried, and disinfected in a hypochlorite sodium solution (0.05%, w/v) for 2 min and rinsed in sterile water and allowed to dry. Sterile pieces were transferred to sterilized Potato Dextrose Agar (PDA) medium to be purified. On PDA solid medium, all fungal colonies were transferred and purified by using single-spore procedures. For subsequent research, purified fungi were stored at 25°C on PDA slants (**sun et al., 2010**).

# 5.3. Effect of SA and Fenhexamid on the linear growth *in vitro*

According to **Townsend and Heuberger (1943),** the warmed, sterilized PDA medium (19 ml) was mixed with one ml of three concentrations of salicylic acid (1, 2 and 4 mM) or Fenhexamid (recommended dose) individually before being poured into plates (9 cm in diameter). Following their solidification, the plates were infected with a disc (5 mm) cut from the peripheral active mycelia development of 5-day-old cultures of a fungal colony and placed in an incubator at 27 °C. Control plates were those that weren't treated. Three plates were employed. All plates' linear fungal growth averages were measured after one week. The mycelia growth inhibition percentage (MGI%), was calculated using the following formula:

#### **MGI% =DC-DT/DC x 100**

Where DC is the control's average mycelia growth's diameter. DT stands for the treatment's average mycelia growth diameter.

# 5.4. Infection % of strawberry fruits during storage under lab condition

SA and Fenhexamid effects on infection rates on strawberry fruits were recorded in the 2020–2021 season at a commercial maturity stage of 3/4 of full fruit color. The identification was confirmed at the Disease Survey and Mycology Department, Plant Pathology Institute, Agriculture Research Center, Egypt. The fungal infection fruits from each treatment were transferred to storage for five days under lab conditions. Then the infection rate of strawberries was calculated according to **Abd-El-Moity**, **1985**) as follows:

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# 5.5. Disease incidence % of gray mold

Plant disease incidence can be defined as the number of plant units that are (visibly) diseased, usually relative to the total number assessed. The effect of SA and Fenhexamid on disease incidence % of strawberry fruits was evaluated according to **Romanazzi** *et al.*, **2000** as follows:

Disease incidence % = [Number of rotted fruits / Total number of tested fruits]  $\times$  100 Analytical statistics





Using the COSTAT package application, the statistical analysis was carried out. An analysis of variance was performed on the data (ANOVA). Using Duncan's Multiple Range Test, the differences between data means were compared (**Waller and Duncan, 1969**). For all statistical calculations, P = 0.05 was used.

#### **Results and Discussion**

1. Effects of SA and Fenhexamid on strawberry vegetative growth and fruit quality

#### **1.1.** Strawberry vegetative growth

In Table 3, the impact of foliar SA treatments on the vegetative growth phenomenon of strawberries in fields with clay and sandy soil is depicted.

**In clay soil field,** application of SA showed no significant effect on leaves number per plant at the rate of 1- and 2-mM recording 7 and 8.3/plant, respectively. Meanwhile, treatments of SA (4 mM) and Fenhexamid significantly increased leaves number/plant being 9.7 and 9.5, respectively, compared to control.

Treatment of SA (1mM), and Fenhexamid, showed no significant effect on flower number / plant or the root length comparing with control, but it showed significant increase when treated with SA at the rate of 2 and 4mM recording 6.5 and 6.5 flowers / plant, or 21.3 and 23 cm root length, respectively.

Treatment of SA at the rate of 4 mM showed the highest significant increase fresh weight of root per plant compared to control followed by the rate of 2 and 1 mM of SA and Fenhexamid recorded 7.2, 5.3, 5.1 and 4.4g, respectively.

**In sandy soil field,** the application of SA at 4mM showed a significant increase in strawberry leaves number (8.3) compared to control (7). Conversely, there was a significant decrease in the leaves number when plants treated with SA (1mM) and Fenhexamid being 3.7 and 3.8, respectively.

The number of flowers per plant did not show significant differences between SA (2 and 4 mM), treatments and control. The plant root length showed no significant differences between SA treatment (2 mM), and control, meanwhile, whereas it recorded the maximum significant increase when applied at the rate of 4mM recording 21.5 cm. Application of Fenhexamid decreased the root length significantly (14cm) compared to control. Also, root fresh weight per plant showed no significant change between Fenhexamid and control. Conversely, SA treatments of 1 and 2 mM increased the root fresh weight significantly compared to control indicating 4.5 and 4.9g, respectively, and reaching its maximum increase when applied at rate of 4 mM (5.7g).

# **1.2.** Fruit quality

**In clay soil field** the highest significant fruit fresh weight was recorded by application of SA at 4mM comparing with the untreated being (30.7g), while there were no significant differences noticed when applied at 1and 2 mM. In contrast, Fenhexamid showed the lower significant fruit fresh weight (15.9g) compared to control. Also, the maximum significant increase in fruit volume was recorded by SA treatment (4mM) compared to control being 40cm<sup>3</sup>, whereas the other treatments showed no significant change.





**In sandy soil field,** the fresh weight of fruit showed no significant changes when treated with 1mM of SA, Fenhexamid, compared to control (19.9, 19.2, and 18.2g, respectively), but it increased significantly and recorded 30.9g when treated with 4mM of SA compared to control. The volume of each fruit showed no significant differences when treated with 1mM of SA, Fenhexamid, comparing with the untreatment, meanwhile, the fruit volume showed significant increase when treated with 4mM of SA being 40 cm<sup>3</sup> compared to control.

These results are in match with **Karlidag** *et al.* (2009), who indicated that salicylic acid dramatically increased strawberry plant growth overall in terms of root length and fresh weight, and it also triggered flowering and fruit setting with higher fruit weight, volume and yield (Fariduddin *et al.*, 2003, Martin-Mex *et al.*, 2005 and Yildirim *et al.*, 2008), According to Lolaei *et al.* (2012), salicylic acid improved fruit output and quality while delaying strawberry ripening. According to AOAC (2005); Mady (2009) and Jamali *et al.* (2011), a salicylic acid treatment at 2.0 mM raised the fresh weights of the roots and shoots, the production of fruit, and the quality of the fruit which three times significantly enhanced the fresh and dry weights of the roots and the rate of vegetative growth when compared to the untreated ones.

# 2. The impact of Fenhexamid and salicylic acid on the pigments of strawberry leaves at two different fields

Table 4 showed the impact of foliar SA application on strawberry leaves pigments at two different fields (clay and sandy soils) at 10, 20, and 30 days after applications.

The data indicated that SA at the rate of 4mM increased all leaves pigments, Chl. A, B, and total chlorophyll, significantly compared to control and the other SA application rates (2 and 1 mM), recorded total chlorophyll value of 3.74 and 3.92 mg/g fw after 10 d., 3.2 and 3.65 mg/g fw after 20 d. and 5.95 and 5.36 mg/g fw after 30 d., from application in clay and sandy soil trial, respectively. Meanwhile, the other 2 applied SA rates (2 and 1 mM), did not show significant effect on the leaves pigments comparing with the untreatment. Conversely, Fenhexamid application showed significant decrease effect on the leaves pigment (Chl. A, B, and total chlorophyll), compared to control after 10, and 20 days after application being 3.18, and 2.36 mg/g fw, and 2.44 and 2.56 mg/g fw of total chlorophyll in the two-trial soil types: clay and sandy soil, respectively. Concerning to Carotenoids leaves content, all treatments did not reveal clear effect on carotenoids content at 10 and 20 days after application compared to control. After 30 days, SA treatment at the rate of 4 mM showed the highest significant increase effect on carotenoids, while Fenhexamid showed the lowest decrease effect being (0.28 and 0.25 mg/g fw), and (0.13 and 0.12 mg/g fw), in clay and sandy soil trial, respectively.

The above-mentioned results are in match with **Arfan** *et al.*, **2007** who indicated that SA increased photosynthetic activity and it is essential to plant growth, ion absorption, and nutrient transport inside the plant. It has been implicated in a crucial function in plant water interactions, photosynthesis, and growth. Also, **Vicente and Plasencia**, **2011** showed that SA is a photosynthesis regulator because of its ability to change the structure of the leaf and chloroplasts. Exogenous administration of SA has an impact on a variety of varied plant activities, including





stomata closure, ion uptake and transport, membrane permeability, photosynthetic and growth rates (Khan and Kenhadji, 2003 and Gunes *et al.*, 2005).

Fenhexamid causes a decline in Pn and in photosynthetic pigment concentrations 7 days after application and decrease in chlorophyll contents and carotenoids after fungicide treatments appeared to be dose-dependent (**Saladin** *et al.*, **2003**).

#### 3. The impact of SA and Fenhexamid on the total phenol in strawberry leaves

Data in Table 5 showed the impact of foliar SA treatments and Fenhexamid on the total phenol of strawberry leaves at two different fields, clay, and sandy soils.

The data indicated that SA treatment had significant increase effect on total phenol content in strawberry leaves at all application rates, in both of clay and sandy soil trial fields meanwhile Fenhexamid treatment showed the lowest significant decrease effect compared to control. The highest phenol content was recorded by SA treatment applied at the rate of 4 mM, being 34.89, 44.23 and 41.96 mg/ml in clay soil trial field, and 29.86, 46.27, and 38.03 mg/ml in sandy soil field after 10, 20 and 30 days from application, respectively.

These results are in agree with **Gad** *et al.*, **2007 and El-Korany and Mohamed**, **2008**, who indicated that salicylic acid resulted in appreciable increases in the total phenol when was sprayed with 4 mM.

# 4. The impact of SA and Fenhexamid on the activity of enzymes

#### 4.1. Catalase activity

Data in Table 6 showed that of SA at all application rates revealed significant increase effect on catalase activity of strawberry leaves at the two tested soil types while application of Fenhexamid showed no significant difference comparing with control. The highest increase effect is recorded by SA at the rate of 4 mM, being 67.1, 94.7 and 160.0 u/mg protein, in clay soil, and 72.4, 107.7, and 180.0 u/mg protein, in sandy soil, after 10, 20, 30 days from application, respectively.

#### 4.2. Phenol oxidase activity

Table 6 illustrated that foliar application of SA at 4 and 2 mM significantly increased phenol oxidase activity compared to the untreatment, meanwhile the treatment of SA at 2mM and Fenhexamid did not indicate significant effect at the two tested soil types after 10, 20, and 30 days comparing with control. The highest effect on phenol oxidase activity is recorded by SA treatment at the rate of 4 mM at 30 days after application being 505.0- and 645.0-units x 1000/min/mg protein in clay and sandy soil, respectively.

#### 4.3. Peroxidase activity

In clay soil field, Table 6 illustrates that all SA treatments revealed a substantial rise in peroxidase at 4, 2, 1 mM application rates compared to control after 10 days from application recorded 564.7, 523.0, and 501.0  $\Delta$ O.Dx1000/min/mg protein, respectively. Meanwhile there was no significant effect at 20 and 30 days from application except the rate of 4 mM which shows the





highest significant rises comparing with control being 650.0 and 846.7  $\Delta$ O.Dx1000/min/mg protein, respectively. In contrast, Fenhexamid showed no significant effect.

In sandy soil, the presented data (Table 6), illustrated that all SA treatments and Fenhexamid showed significant effect on peroxidase activity after 20 and 30 days from treatment. The highest rise effect was recorded by SA (4mM), being 11053 and 1211.7  $\Delta$ O.Dx1000/min/mg protein, after 20 and 30 days,

The results of strawberry enzymes activity agree with **Khan** *et al.*, **2012**; **and Shafiee**, *et al.*, **2010** who found that SA has been demonstrated to be a significant signal molecule that can activate a specific enzyme that catalysis biosynthetic reaction. Exogenous SA has been applied to plants in recent years at non-toxic concentrations to control a variety of processes, including biotic and abiotic stressors. Also, **Hayat** *et al.* (**2010**) indicated that SA an endogenous plant hormone is produce antioxidant enzymes.

#### 5. Impact of SA and Fenhexamid on fungus

#### 5.1. Effectiveness on mycelia growth inhibition in vitro.

Data indicated in figure 1 revealed that SA treatments showed a promising inhibition effect on *B. cinerea* MGI %. Fenhexamid fungicide and SA showed positive linear inhibition effect to the mycelia growth by increasing application rate. Salicylic acid treatment showed close inhibition effect to Fenhexamid at the lower tested rate of 200, 400, and 600 ppm being (51.3 and 55.4%), (71.5 and 80.7%) or (87.1 and 89.5%), respectively. At the higher application rate (800 ppm), each of SA and Fenhexamid showed the same inhibition effect recoding 94.4%.

# 5.2. Salicylic Acid and Fenhexamid effects on strawberry fruit gray mold when being stored under laboratory conditions.

Results in Table 7 demonstrated the infection % of strawberry fruits collected from clay and sandy soil trial, during storage under laboratory conditions. Gray mold infection % of SA at the different application rates and Fenhexamid were markedly lower than control. According to the data, SA at the rate of 1 mM, and Fenhexamid treatment (at the recommended dose) showed equivalent effect and reduced the infection % by 21.4, and 19.4% or 21.4 and 18.6% of clay and sandy soil trial, respectively. Application of SA at 4 mM showed high positive effect and reduced the infection % more than Fenhexamid fungicide being 10.9 and 19.5% or 10.9 and 18.7% for clay and sandy soil trial, respectively.

#### 5.3. Effectives of SA and Fenhexamid on disease incidence%

Data in Table 8 indicated the effect of SA and Fenhexamid on the disease incidence % of strawberry gray mold under field conditions.

The data showed that the incidence of strawberry fruit gray mold in SA and Fenhexamid treatments was markedly lower than the untreatment. Treatment of SA at 1 mM showed a slight difference effect comparing to Fenhexamid fungicide, being 27.3 and 24.6 % or 26.7 and 23.4 % of clay and sandy soil trial, respectively. At the higher application rates of SA (2 and 4 mM), were





markedly reduced the infection % compared to Fenhexamid treatment, recorded (15.2 and 4.3%) or (24.6 and 24.7%) in clay soil, or (15.0 and 4.2%) or (23.6 and 23.6%) in sandy soil, respectively.

These results are in match with Ezzatet al., 2017 who indicated that the application of salicylic acid to strawberry seedlings resulted in much less gray mold on the seedlings and improved seedling development as compared to the control. Also, phenolic compounds will protect seedlings by enhancing their disease resistance, promoting seedling growth, increasing their tolerance to environmental challenges, and improving plant growth and productivity. Also, it could be due that SA treatment efficiently lower fruit ethylene production, inhibit fungal rot, and maintains overall fruit quality (El-Morsy et al., 2022). Also, SA postharvest treatment of fruit inhibits the growth and spread of rot fungi in strawberry fruits (Doares et al. (1995); Huang et al. (2000); Amborabe et al., (2002); Lu and Chen (2005); Babalar et al. (2007) and Vicente and Plasencia, 2011).

# References

**Abd-El-Moity, T.H. (1985).** Effect of single and mixture of *Trichodermaharzianum* isolates on controlling three different soil-borne pathogens. Egypt. J. Micro., 111-120.

Aebi, H. (1984). Catalase in vitro. Methods in enzymology. 105:121-126.

Amborabe, B. E., Lessard, P. F., Chollet, J. F., and Roblin, G. (2002). Antifungal effects of salicylic acid and other benzoic acid derivatives towards Eutypalata: Structure–activity relationship. Plant Physiology and Biochemistry, 40, 1051–1060.

Arfan, M.; Athar, H.R. and Ashraf, M. (2007). Does exogenous application of salicylic acid through the rotting medium modulate growth and photosynthetic capacity in two differently adapted spring wheat cultivars under salt? J. plant physiology, 164(6): 685-694.

Arnon, D.I. (1949). Copper enzymes in isolated chloroplasts, poly phenol oxidase in Beta vulgaris. Plant Physiol. 24 (1): 1-15.

Association of Official Analytical Chemists-International. (2005). Official Methods of Analysis. 18<sup>th</sup> edn., eds.: W. Hortwitz, G. W. Latimer, AOAC-Int. Suite 500, 481 North Frederick Avenue, Gaithersburg, Maryland, USA.

**Babalar, M., M. Asghari, A. Talaei and A. Khosroshahi.** (2007). Effect of pre- and postharvest salicylic acid treatment on ethylene production, fungal decay and overall quality of Selva strawberry fruit. Food Chemistry, 105: 449-453.

**Bradford, M.M.** (1976). A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. Analytical Biochemistry, 72: 248-254.

**Cañal, V. M. J.; Fernandez, B. M. and Sanchez, R. T. (1985).** Effect of glyphosate on growth and the chlorophyll and carotenoid levels of yellow Nutsedge (*Cyperusesculentus*). Weed Sci., 33: 751-754.





**Chauhan, S.K.; Dhillon, W.S.; Singh, N. and Sharma, R. (2013).** Physiological behavior and yield evaluation of agronomic crops under agri-horti-silviculture system. Inter. J. plant Res. 3(1):1-8.

**Ciliberti, N.; Fermaud, M.; Roudet, J. and Rossi, V. (2015).** Environmental Conditions Affect *Botrytis cinerea* Infection of Mature Grape Berries More Than the Strain or Transposon Genotype. Phytopathology. 105 (8): 1090–1096.

**Deising, H.B.; Reimann, S. and Pascholati, S.F. (2008).** Mechanisms and significance of fungicide resistance. Braz. J. Microbiol., 39(2): 286–295.

**Doares, S.H.; Narvaez-Vasquez, J. Conconi, A. and Ryan, C. A. (1995).** Salicylic acid inhibits synthesis of proteinase inhibitors in tomato leaves induced by systemin and jasmonic acid. Plant physiology, 108(4): 1741-1746.

**El-falleh W, Hannachi H, Tlili N, Yahia Y, Nasri N, Ferchichi A (2012).** Total phenolic contents and antioxidant activities of pomegranate peel, seed, leaf and flower. J. Med. Plants Res., 6: 4724-4730.

**El-Korany, A.E. and Mohamed, R.A. (2008)**. The use of antioxidants to control gray mold and to enhance yield and quality of strawberry. J. Agric. Env. Sci. Alex. Univ. Egypt. 7(1):1-38.

**El-Morsy, M. M.; Koriem, A. M.; Khafagi, E. Y. and Elian, M. I. (2022).** Controlling strawberry fruit rots with the alternative of fungicides. Inter. J. Scientific Res. and Sustainable Develop. 5(2):1-12.

Ezzat, A.; Ammar, A.; Szabó, Z.; Nyéki, J. and Holb, I.J. (2017). Postharvest Treatments with Methyl Jasmonate and Salicylic Acid for Maintaining Physico-Chemical Characteristics and Sensory Quality Properties of Apricot Fruit during Cold.

**Fariduddin, Q.; Hayat, S. and Ahmad, A. (2003).** Salicylic acid influences net photosynthetic rate, carboxylation efficiency, nitrate reductase activity and seed yield in Brassica juncea. Photosynthetica, 41: 281–284.

Gad, N., M.H. Elsherif, and N.H.M. Elgereed. (2007). Influence of nickel on some physiological aspects of tomato plants. Australian J. Basic Appl. Sci. 1 (3):286-293.

Gunes, A.; Inal, A.; Alpaslan, M.; Cicek, N.; Guneri, E. and Eraslan, F. (2005). Effects of exogenously applied salicylic acid on the induction of multiple stress tolerance and mineral nutrition in maize (*Zea mays* L.) (EinflusseinerSalicylsaure-Applikation). Archives of agronomy and soil Sci. 51(6):687-695.

Ha nßler. G. and Pontzen, R. (1999). Effect of fenhexamid on the development of *Botrytis cinerea*. Pflanzen Nach Bayer, 52:158–176

Hammerschmodt, R.; Nuckles, E.M. and Kuc, J. (1982). Systemic resistance of cucumber to *Colletotrichumlagenarium*. Physiological plant Pathology, 20(1):73-82.

Hamouda, H. A1.; Khalifa, R. Kh. M.; El-Dahshouri, M. F. and Nagwa G. Zahran (2016). Yield, fruit quality and nutrients content of pomegranate leaves and fruit as influenced by iron, manganese, and zinc foliar spray. International Journal of Pharm Tech Research, 9(3):46-57.

Hayat, Q.; Hayat, S.; Irfan, M. and Ahmad, A. (2010). Effect of exogenous salicylic acid under changing environment: A review. Environmental and Experimental Botany 6: 14-25.





Hiscox, J. D. and Israelstam, G. F. (1979). A method for the extraction of chlorophyll from leaf tissue without maceration. Can. J. Bot. 57: 1332-1334.

Huang, Y.; Deverall, B. J; Tang, W.H; Wang, W., and Wu, F.W. (2000). Foliar application of acibenzolar-S-methyl and protection of postharvest rock melons and Ham melons from disease. European J. plant Pathology, 106(7): 651-656.

Hull, H.M.; Morton, H.L. and Wharrie, J.R. (1975). The Botanical Rev. 41(4): 421-452.
 Ishaaya, I. (1971). Observations on the phenol oxidase system in the armored scales
 Aonidiellaaurantii and Chrysomphalusaonidum.Comparative Biochem. And Physiol. Part B:
 Comparative Biochem. 39(4): 935-943.

Jamali, B, Eshghi, S. and Tafazoli, E. (2011). Vegetative and reproductive growth of strawberry plants cv. Pajaro by salicylic acid and nickel. J. Agric. Sci. Tech.; 13: 895-904.

Karlidag, H.; Yildirim, E. and Turan, M. (2009). Salicylic acid ameliorates the adverse effect of salt stress on strawberry. Sci. Agric. 66: 180-187.

Khan, M. I.R.; Syeed, S.; Nazar, R. and Anjum, N.A. (2012). An insight into the role of salicylic acid and jasmonic acid in salt stress tolerance. Phytohormones and abiotic stress tolerance in plants, 277-300.

Khan, M.S. and Kenhadji, A. (2003). Financial development and economic growth: A review and new evidence. J. African Economies, 12: 89-110.

Kim, J.H.; Lee S.H.; Kim, C.S.; Lim, E.K.; Choi, K.H.; Kong, H.G.; Kim, D.W.; Lee, S.W.; Moon, B.J. (2007). Biological control of strawberry gray mold caused by *Botrytis cinerea* using *Bacillus licheniformis* N1 formulation. J. Microbiology Biotechnology. 17(3): 438-444.

Lolaei, A.; Kaviani, B.; Rezaei, M.A. and Raad, M.K. (2012). Effect of pre-and postharvest treatment of salicylic acid on ripening of fruit and overall quality of strawberry (*Fragariaananasa*Duch cv. Camarosa) fruit. Annals if Biological Res.3(10): 4680-4684.

Lu, Y. Y., and Chen, C. Y. (2005). Molecular analysis of lily leaves in response to salicylic acid effective towards protection against Botrytis elliptica. Plant Science, 169, 1–9.

**Mady, MA. (2009).** Effect of foliar application with salicylic acid and vitamin E on growth and productivity of Tomato (*Lycopersicon Esculrntum*, Mill.) plant. J. plant production 34(6): 6715-6726.

Martin-Mex, R., E. Villanueva, T, Herrera, and Larque-Saaverda. (2005). Positive effect of salicylates on the flowering of cucumber. Aus. J. Bas. Appl. Sci. 2(11):879-883.

Meyers, K.J.; Watkins, C.B.; Pritts, M. P. and Liu, R.H. (2003). Antioxidant and antiproliferative activities of strawberries J. Agric. Food Chem. 5(23): 6887-6892.

Ni, L.; Jie, X.; Wang, P., Li, S.; Wang, G. Li, Y. and Li, Y. (2015). Effect of linoleic acid sustained-release microspheres on *Microcystis aeruginosa* antioxidant enzymes activity and microcystins production and release Chemosphere. 121:110-116.

**Peng, L. and Jiang, Y. (2006).** Exogenous salicylic acid inhibits browning of fresh-cut Chinese water chestnut. Food Chem., 94: 535-540.

Raskin, I. (1992). Role of salicylic acid in plants. Ann. Rev. plant Bio. 43(1): 439-463.





Romanazzi, G., Nigro, F., Ippolito, A. (2000). Effectiveness of pre and postharvest chitosan treatments on storage decay of strawberries. Frutticoltura, 62: 71–75.

Saladin, G.; Magne, C. and Cle´ment, C. (2003). Effects of fludioxonil and pyrimethanil, two fungicides used against *Botrytis cinerea*, on carbohydrate physiology in *Vitis vinifera* L. Pest Manag. Sci. 59:1083–1092.

Shafiee, M.; Taghavi, T.S. and Babalar, M. (2010). Addition of salicylic acid to nutrient solution combined with postharvest treatments (hot water, salicylic acid, and calcium dipping) improved postharvest fruit quality of strawberry. ScientiaHorticulturae, 124: 40-45.

Sun H. Y., Wang H. C., Chen Y., Li H. X., Chen C. J., Zhou M. G. (2010). Multiple resistance of *Botrytis cinerea* from vegetable crops to carbendazim, diethofencarb, procymidone, and pyrimethanil in China. *Plant Disease*, 94: 551–556.

**Sutton, J.C. (1990).** Epidemiology and management of Botrytis leaf blight of onion and gray mold of strawberry: a comparative analysis. Canadian J. plant Pathology. 12(1): 100-110.

Sutton, J.C. and Peng, G. (1993). Biocontrol of *Botrytis cinerea* in strawberry leaves. Phytopathology. 83(6):615-621.

Townsend, G.R. and Heuberger, J.W. (1943). Methods for estimating losses caused by diseases in fungicide experiments. The Plant Disease Reporter, 27: 340-343.

Vicente, M.R.S. and Plasencia, J. (2011). Salicylic acid beyond defense: its role in plant growth and development. Journal of Experimental Botany: 1-18.

**Waller, RA and Duncan, DB. (1969).** A Bayes rule for the symmetric multiple comparisons problem. J. American Statistical Association, 64(328): 1484-1503.

**Yildirim, E and Dursun, A. (2008).** Effect of foliar salicylic acid applications on plant growth and yield of tomato under greenhouse conditions. Inter. Symposium on strategies towards sustainability of protected cultivation in mild winter climate, 807:395-400.

Zhang, F.; Zhang, H.; Xia, Y. Wang, G.; Xu, L. and Shen, Z. (2011). Exogenous application of salicylic acid alleviates cadmium toxicity and reduces hydrogen peroxide accumulation in root apoplasts of *Phaseolus aureus* and *Vicia sativa*. Plant cell Reports 30(8): 1475-1483





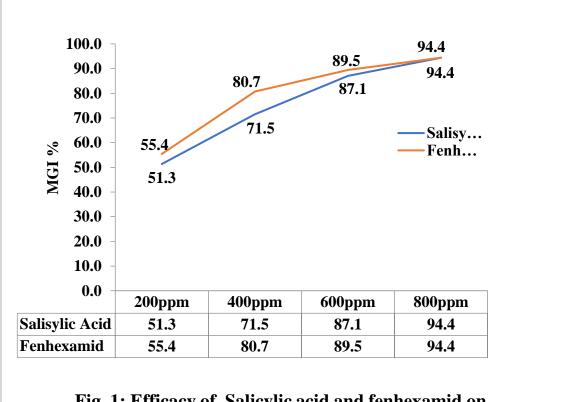


Fig. 1: Efficacy of Salicylic acid and fenhexamid on mycelia growth inhibition of strawberry fruits rot caused by *Botrytis Cinerea in vitro* 





		Ι	Physical ]	properties				SP%		EC			ոՍ	
	Clay%	Silt	:%	Sand%	Total%	Texture	;	SF 70		DS/m	ı		рН	
	6	8	3	86	100	Clay		50		0.18			8.3	
					Ch	emical pro	operties							
S	Soluble ca	tions (meq	[/L)	1	Soluble anions	(meq/L)			]	Mineral	element	ts (ppm)		
Ca <sup>++</sup>	$\mathbf{Mg}^{++}$	$Na^+$	$\mathbf{K}^{+}$	$CO_{3}^{-2}$	$HCO_3^{-1}$ (	CI <sup>-1</sup> S	504 <sup>-2</sup>	Ν	Р	К	Cu	Fe	Mn	Zn
0.8	0.4	0.4	0.2	-	0.2	1	0.6	30	42	128	1.3	25.68	-	0.58
able 2	2: Chemic	al and ph	nysical p	roperties	of the expren	nintal fiel	d locati	ion B						
able	2: Chemio	•		roperties properties	•	nintal field	d locati			EC				
	2: Chemic	•	Physical	•	•			ion B SP%		EC DS/1			рН	
		Sil	Physical	properties	5						m		<b>pH</b> 8.2	
	Clay%	Sil	Physical t%	properties Sand%	<b>Total%</b> 100	Textur	e	<b>SP%</b> 50		DS/1	m			
	<b>Clay%</b> 10.88	Sil	Physical t% 0	properties Sand% 79.12	<b>Total%</b> 100	Textur Sand nemical pr	e	<b>SP%</b> 50		<b>DS</b> /1 0.5	<b>m</b> ;	nts (ppm)	8.2	
	<b>Clay%</b> 10.88	] Sil 1	Physical t% 0	properties Sand% 79.12	Total% 100 Cł Soluble anions	Textur Sand nemical pr s (meq/L)	e	<b>SP%</b> 50	Р	<b>DS</b> /1 0.5	<b>m</b> ;	nts (ppm) Fe	8.2	Zn

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# Table (3) Effect of salicylic acid applications on Strawberry vegetative growth phenomenon at two different soil typesof fields at 10, 20 and 30 days after treatment.

						Cla	ıy so	oil field	1									Sa	ındy	soil fie	eld				
Treatment	Appl. Rate	l. Rate Leaves No./ Plant				Root L/ Plant (Cm)		Root FW/ Plant (g)		Fruit FW g/ fruit		Frui Vol (cm <sup>3</sup> ) fruit	)/	Leaves No./ Plant		Flower No./ Plant		Root L/ Plant (Cm) Root FV Plant (§					Fruit (cm fru	<sup>3</sup> )/	
ic	4 mM	9.7	а	6.5	a	23.0	a	7.2	a	30.7	Α	40.0	а	8.3	a	6.2	а	21.5	a	5.7	а	30.9	a	40.0	а
Salicylic Acid	2 mM	8.3	ab	6.5	a	21.3	a	5.3	b	22.2	В	30.0	b	6.5	b	5.7	а	19.0	b	4.9	ab	25.1	b	30.0	b
Sa	1 mM	7.0	bc	4.0	b	17.0	b	5.1	b	19.5	С	20.0	c	6.3	b	3.7	с	16.3	с	4.5	b	19.9	c	20.3	c
Un-Treat.		6.3	с	3.3	b	16.7	b	2.7	с	18.5	С	20.3	c	7.0	b	5.0	ab	18.7	b	2.7	с	18.2	c	21.0	c
Fenhexamid SC 50%	600 CC/ Fed.	9.5	a	4.5	b	14.3	b	4.4	b	15.9	D	19.7	c	6.8	b	3.8	bc	14.0	d	2.8	с	19.2	с	20.0	с
LSD (0.05) 1.467 1.834		34	4.08	4.088 0.830		80	1.229		1.558		1.151		1.243		1.97	.979 0.81		10	1.759		2.048				

Same letter at same column is not significant.





# Table (4) Effect of salicylic acid applications on Strawberry leaves pigments (mg/g fresh weight) at two different soil types of fields at 10,20 and 30 after treatment.

										10	) Da	ays								
Treatments	Appl. Rate				C	lay soil fiel	ld									Sandy soil f	field			
		Caroter	ne	Chl. A	1	Chl. B		Total C	hl.	A/B ratio		Caroten	e	Chl. A	1	Chl. 1	В	Total Chl.		A/B ratio
Salicylic Acid	4 mM	0.27	а	2.59	а	1.15	а	3.74	Α	2.25		0.28	a	2.60	а	1.32	а	3.92	а	1.97
licy	2 mM	0.23	b	2.35	ab	1.05	b	3.39	В	2.24		0.19	b	2.18	b	1.03	b	3.21	b	2.19
Sa] ≁	1 mM	0.23	b	1.91	с	1.04	b	2.95	С	1.84		0.18	b	1.91	с	0.94	bc	2.85	с	2.04
Un-Treat		0.27	а	2.18	b	1.14	а	3.32	В	1.91		0.27	a	2.40	ab	1.01	b	3.41	b	2.38
Fenhexamid SC 50%	600 CC/ Fed.	0.23	b	2.16	b	1.02	b	3.18	Bc	1.79		0.18	b	1.60	d	0.78	с	2.38	d	2.06
LSE	)	0.0205		0.2451		0.0477		0.2600				0.0290		0.2544		0.1902		0.3483		
Treatment	Appl. Rate								20	) Da	ays				-					
	Аррі. Кас	Carotene		Chl. A		Chl. B		Total C	hl.	A/B ratio		Caroten	e	Chl. A	1	Chl. 1	В	Total C	hl.	A/B ratio
Salicylic Acid	4 mM	0.17	а	2.65	а	0.64	а	3.29	Α	4.12		0.19	a	2.99	а	0.66	а	3.65	а	4.53
Aci	2 mM	0.15	ab	2.40	ab	0.54	b	2.94	Bc	4.47		0.17	a	2.44	b	0.63	ab	3.07	bc	3.91
Sa	1 mM	0.13	b	2.29	b	0.53	b	2.82	С	4.32		0.13	b	2.25	b	0.57	bc	2.81	cd	4.03
Un-Treat.		0.16	ab	2.62	а	0.52	b	3.14	Ab	5.03		0.18	a	2.59	b	0.62	abc	3.22	b	4.15
Fenhexamid SC 50%	600 CC/ Fed.	0.14	ab	2.00	с	0.44	c	2.44	D	4.55		0.12	b	2.02	с	0.54	с	2.56	d	3.72
LSE	)	0.0282		0.2467		0.0469		0.2504				0.0300		0.3310		0.0778		0.3278		
Treatment	Appl. Rate										Da	ays								
	Аррі. Кас	Caroter	ne	Chl. A	4	Chl. B		Total C	hl.	A/B ratio		Caroten	e	Chl. A	1	Chl. I	В	Total C	hl.	A/B ratio
d d	4 mM	0.28	а	4.73	а	1.23	а	5.95	а	3.86		0.25	a	4.33	а	1.04	а	5.36	а	4.18
Salicylic Acid	2 mM	0.23	b	3.93	b	0.75	с	4.69	b	5.22		0.22	b	4.22	а	0.97	ab	5.19	а	4.35
	1 mM	0.23	b	2.85	с	0.92	b	3.77	с	3.11		0.21	b	2.78	с	0.94	ab	3.72	с	2.96
Un-Treat.		0.22	b	4.02	b	0.96	b	4.98	b	4.19		0.22	b	3.45	b	1.01	а	4.45	b	3.42
Fenhexamid SC 50%	600 CC/ Fed.	0.13	с	3.68	b	0.93	b	4.60	b	3.99		0.12	c	3.36	b	0.88	b	4.24	b	3.87
LSD (0	.05)	0.0194		0.3465		0.1082		0.4311				0.1999		0.2424		0.1084		0.2470		

Same letter at same column is not significant.





#### Table (5) Effect of salicylic acid applications on total phenol contents (mg/ml) of Strawberry leaves at two different soil types of fields at 10, 20 and 30 days after treatment.

Same at					Clay soil fi	eld			Sandy soil field						
at	Treatment	Appl. Rate	10 d		20 d		30 d		10 d		20 d		30 d		s
	Acid	4 mM	34.89	a	44.23	a	41.96	a	29.86	a	46.27	a	38.03	a	
	Salicylic ,	2 mM	33.72	b	36.69	b	40.16	b	28.73	b	44.47	b	36.03	b	
	Salic	1 mM	24.83	с	31.42	с	34.71	с	26.59	c	39.90	с	33.10	c	
	Un-Treat.		23.42	d	22.11	d	28.96	d	22.44	d	31.96	d	32.56	с	]
	Fenhexamid600 CC/SC 50%Fed.		21.84	e	21.80	e	23.83	e	18.87	e	25.13	e	25.76	d	
	LSD (	0.855		0.815		0.626		0.412		0.603		1.096		1	

column is not significant.





Table (6) Effect of salicylic acid and fenhexamid on Strawberry leaves enzymes at two different soil types of fields at 10, 20 and 30 days
after treatment.

						aller treatine	/110.						
	Turaturata			Clay soil	field					Sandy soil fi	eld		
(u)	Treatments	10 d		20 d		30 d		10 d		20 d		30 d	
otei	SA-4mM	67.1	a	94.7	a	160.0	a	72.4	a	107.7	a	180.0	a
atalase ng pro	SA-2mM	60.8	b	86.0	b	125.0	b	62.2	b	92.0	b	142.3	b
Catalase (unit/mg protein)	SA-1mM	53.0	с	72.0	с	117.7	bc	56.7	с	82.0	с	132.0	bc
uit/1	Fenhexamid	53.2	с	68.0	cd	113.7	с	53.1	d	74.0	d	122.7	с
(n	control	51.8	с	64.0	d	117.0	bc	52.9	d	69.0	e	116.7	с
	LSD	2.2537		4.6973		9.718		2.3407		3.5773		16.93	
e e	SA-4mM	386.0	a	437.0	а	505.0	a	404.0	а	445.0	а	645.0	а
dase /mg )	SA-2mM	369.3	b	431.7	a	483.3	b	388.0	b	432.7	b	550.0	b
oxidase inits /min/mg (tein)	SA-1mM	364.7	bc	422.0	b	462.0	d	372.0	с	428.0	b	506.7	с
	Fenhexamid	364.3	bc	418.0	bc	468.0	С	368.0	cd	421.0	с	511.7	с
Phenol (U x1000/ pro	control	360.7	с	412.7	с	461.0	d	366.0	d	405.3	d	505.3	с
P] x	LSD	6.7256		5.9602		5.6563		5.5181		6.2317		11.6774	
e /mg	SA-4mM	564.7	a	650.0	а	846.7	a	1035.0	а	1105.3	а	1211.7	а
lse n/n	SA-2mM	523.0	b	620.0	b	725.0	b	907.3	b	1002.0	b	1180.0	b
iida O. ein	SA-1mM	501.0	с	594.7	с	720.0	b	710.0	с	801.0	с	975.0	с
Peroxidase (∆O. 1000/min/1 protein)	Fenhexamid	485.0	d	591.0	с	722.3	b	663.3	cd	760.0	d	915.7	d
Ре (10 р	control	481.7	d	584.7	с	715.0	b	620.0	d	706.0	e	850.0	e
Ŋ	LSD (0.05)	7.4863		12.49		94.5833		51.1319		11.6015		15.4299	

Same letter at same column is not significant.





 Table (7): Efficacy of pre-harvest application of salicylic acid and fenhexamid on gray mold infection % of strawberry fruits during storage under laboratory conditions.

		Clay soil		Sandy soil					
Concentrations	1mM	2mM	4mM		1mM	2mM	4mM		
Salicylic Acid	21.4	13.2	10.9		21.4	13.3	10.9		
Fenhexamid (Recom. dose)	19.4	19.5	19.5		18.6	18.5	18.7		
Control	50.2	50.2	50.2		50.2	50.2	50.2		

Table (8): Efficacy of salicyl	ic acid and fungi	cide treatments	s on gray mold i	infection % unde	r field condition	ns.				
	Clay soil Sandy soil									
Concentrations	1mM	2mM	4mM	1mM	2mM	4mM				
Salicylic Acid	27.3	15.2	4.3	26.7	15.0	4.2				
Fenhexamid (Recom. dose)	24.6	24.6	24.7	23.4	23.6	23.6				
control	48.6	48.6	48.6	48.6	48.6	48.6				

